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Thermodynamic basis for evolution of apatite in calcified tissues

¹⁰ Sabrina Rollin-Martinet^{a,b}, Alexandra Navrotsky^c,

¹¹ Eric Champion^b, David Grossin^a, and Christophe Drouet^{a,*}

¹⁴ ^aCIRIMAT Carnot Institute, University of Toulouse, UMR 5085 CNRS/INPT/UPS, ENSIACET, 4 allee Emile Monso, 31030 Toulouse cedex 4, France (Corresponding author, E-mail: <u>christophe.drouet@ensiacet.fr</u>

¹⁹ ^bUniversité de Limoges, CNRS, SPCTS, UMR 7315, Centre Européen de la Céramique,

20 12 rue Atlantis, 87068 Limoges cedex, France (E-mail: eric.champion@unilim.fr)

²¹ ²² ^cPeter A. Rock Thermochemistry Laboratory and NEAT ORU, University of California Davis, 1 Shields Ave., Davis CA 95616 USA (E-mail: anavrotsky@ucdavis.edu)

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²⁸ Main corresponding Author:

29 Dr. Christophe Drouet

- 30 CIRIMAT Carnot Institute
- 31 ENSIACET
- 32 4 allee Emile Monso
- ³³ 31030 Toulouse cedex 4, France

³⁴ E-mail: christophe.drouet@ensiacet.fr

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40 Abstract

⁴¹ Bone remodeling and tooth enamel maturation are biological processes which alter the physico-42 chemical features of biominerals with time. However, although the ubiquity of bone remodeling is 43 clear, why is well crystallized bone mineral systematically replaced by immature nanocrystalline 44 inorganic material? In enamel, a clear evolution is also seen from the first mineral formed during the 45 secretory stage and its mature well crystalline form, which then changes little in the adult tooth. This 46 contribution provides the thermodynamic basis underlying these biological phenomena. We 47 determined, for the first time, the energetics of biomimetic apatites corresponding to an increasing ₄₈ degree of maturation. Our data point out the progressive evolution of the enthalpy (ΔH_{f}°) and free ⁴⁹ energy (ΔG_f°) of formation toward more negative values upon maturation. Entropy contributions to $_{50}\Delta G_{f}^{\circ}$ values remained small compared to enthalpy contributions. ΔH_{f}° varied from -12058.9 ± 12.2 to $_{\text{s1}}$ -12771.0 ± 21.4 kJ/mol for maturation times increasing from 20 min to 3 weeks, approaching the value s2 for stoichiometric hydroxyapatite, -13431.0 ± 22.7 kJ/mol. Apatite thermodynamic stability increased s as its composition moved toward stoichiometry. These findings imply diminishing aqueous solubility set of calcium and phosphate ions as well as decreased surface reactivity. Such thermodynamically-driven 55 maturation is favorable for enamel maturation since this biomineral is intended to resist external 56 aggressions such as contact with acids. In contrast, maintaining a metastable highly reactive and s7 soluble form of apatite is essential to the effective participation of bone as a source of calcium and s phosphate for homeostasis. Therefore our data strongly suggest that, far from being trivial, the intrinsic ³⁹ thermodynamic properties of apatite mineral represent a critical driving force for continuous bone ⁶⁰ remodeling, in contrast to current views favoring a purely biologically driven cycle. These 61 thermodynamic data may prove helpful in other domains relating, for example, to apatite-based 62 biomaterials development or in the field of (geo)microbiology.

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65 **1. Introduction**

⁶⁶ Calcified tissues are complex adaptive biomaterials optimized through evolution to provide a union of er inorganic and organic constituents to serve both mechanical and biological functions. In particular, one ⁶⁸ can underline the sophisticated multi-scale architectures found in tooth enamel and in bone, which 69 control their mechanical and chemical properties (Weiner and Wagner 1998, Gomez-Morales et al. ⁷⁰ 2013). Along with morphological aspects, the chemical composition, crystal structure and n microstructure of such apatitic biominerals are oadapted to their physiological functions. Enamel, for ⁷² instance, is intended to protect erupted teeth against external aggressions (thermal, mechanical, ⁷³ chemical), and thus requires in its mature state a high degree of chemical and mechanical stability and ⁷⁴ low aqueous solubility. These conditions are met thanks to a chemical composition and other physico-⁷⁵ chemical features for mature enamel close to stoichiometric hydroxyapatite (HA, hexagonal, P6₃/m ⁷⁶ space group) (Bonar et al. 1991). In contrast, far from being inert, bone acts as an ion reservoir τ allowing for the continual regulation of mineral ion concentrations in body fluids (homeostasis) ⁷⁸ (Driessens et al. 1986). Therefore, bone mineral should be relatively soluble and should remain highly ⁷⁹ reactive. Such an increase in solubility and reactivity relative to well-crystallized stoichiometric ⁸⁰ hydroxyapatite can be attained through nonstoichiometry (ion vacancies) as well as by nanometric ⁸¹ crystal dimensions and a low degree of crystallinity (Grynpas 1976).

Without taking into account the presence of secondary elements, the overall composition of biomimetic apatite can generally be satisfactorily described by formulas such as: $Ca_{10-x}(PO_4)_{6-x}(HPO_4)_x(OH)_{2-x}$ (proposed by Winand 1961) or $Ca_{10-x-Z}(PO_4)_{6-x}(HPO_4)_x(OH)_{2-x-2Z}$ (proposed by Kühl and Nebergall 1963), where x and Z depend on factors such as conditions of formation and/or state of ageing. The presence of carbonate ions is also observed, especially in mature biominerals (Gomez-Morales et al. 2013), whereas significantly lower carbonate amounts are found in immature ones (Rey et al. 1995). The reactivity of such nanocrystalline apatites is directly connected to specific substructural features: detailed works on such nanocrystalline apatite compounds, mostly based on spectroscopic studies (Roufosse et al. 1984; Rey et al. 1989a; Rey et al. 1990; Lu et al., 2000; Kaflak et al. 2008), reveal the presence of non-apatitic ionic environments located within a calcium phosphate hydrated layer on the surface of the nanocrystals, whether of synthetic or of biological origin (Cazalbou et al. 2004a; Rey et al. 1989b). This type of complex substructure can probably be related to the mode of formation of apatites (Cazalbou et al. 2004b), which enclose a large number of ions per unit formula and for which the kinetics of crystallization is slow.

The surface layer has been shown to be mostly composed of divalent ions (e.g. Ca²⁺, HPO₄²⁻...), that are rather labile and can be easily and rapidly exchanged (within a few minutes) by other ions from the surrounding fluid (Eichert et al. 2008). Ion exchange isotherms with Langmuir-like features are generally seen (Drouet et al. 2008). Also, the presence of this layer plays a key role in the adsorption of (bio)molecules (Ouizat et al. 1999), and such adsorption phenomena sometimes involve a simultaneous release of surface ions (Errassifi et al. 2010). The presence of this layer generally leads to amorphous-like features on electron microscopy analyses (Sakhno et al. 2010), and solid state NMR data also distinguish between the less-ordered surface ionic environments and bulk species (Wu et al. 2002; Jager et al. 2006; Kaflak et al. 2008).

The preparation of synthetic analogs to biological apatites – i.e. mimicking at the same time compositional, crystallographic and microstructural features – has been made possible at the laboratory rescale using "mild" synthetic conditions, generally through precipitation at room temperature and physiological pH (Rey et al. 1989b; Cazalbou et al. 2004a; Cazalbou et al. 2004b; Drouet et al. 2009). Variations in synthesis protocols, especially modification of temperature, pH and/or maturation time prior to precipitate filtration, significantly alter the structural and chemical characteristics of the nanocrystals (Vandecandelaere et al. 2012). An increase of maturation time (ageing in solution) leads to an increase in mean crystallite dimensions as well as a progressive evolution of the chemical

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¹¹³ composition toward the stoichiometry of hydroxyapatite (Ca₁₀(PO₄)₆(OH)₂, denoted "HA") (Cazalbou ¹¹⁴ et al. 2004a), much like what is observed *in vivo* for enamel maturation between the initial stages of ¹¹⁵ formation (during the secretory stage by ameloblast cells) and its mature state (Rey et al. 1995; ¹¹⁶ Gomez-Morales et al. 2013). Also, a decrease in surface reactivity, measured either by ion exchange or ¹¹⁷ adsorption, has been noticed for crystals matured for increasing periods of time in an aqueous medium ¹¹⁸ (Ouizat et al. 1999; Eichert et al. 2008). In the case of bone, several works have reported the ¹¹⁹ modification in mineral composition and in the amount of non-apatitic surface species upon ageing, ¹²⁰ with an increase of the amount of carbonate and a decrease of the HPO₄ content (Kühl and Nebergall ¹²¹ 1963; Legros et al. 1987; Rey et al. 1991a; Rey et al. 1995). During this ageing process, some ions of ¹²² the surface layer are likely to be incorporated into the apatitic core of the nanocrystals and the overall ¹²³ surface area decreases. These changes may then limit ion exchange which plays a key role in ¹²⁴ physiological pathways (Neuman et al. 1956; Pak et al. 1967; Neuman et al. 1968; Johnson et al. 1970; ¹²⁵ Fernandez-Gavarron 1978; Neuman and Neuman 1985).

Although apatite nanocrystal maturation/ageing has been the object of much investigation, no quantification of the energetics of biomimetic apatite compounds nor of their ageing can be found in the literature to the best of our knowledge; as only thermodynamic data for stoichiometric coarsely rzy crystalline apatitic compounds are available (Jemal et al. 1995; Jemal 2004; Ben Cherifa and Jemal 2004). The object of this contribution is to investigate the energetic evolution of precipitated biomimetic apatite during ageing, based on solution calorimetric studies coupled with careful characterization and chemical analyses, and to relate the energetics to the behavior of apatite nanocrystal biominerals, linked in particular to enamel maturation and to bone remodeling. This study was carried out on non-carbonated apatites, thus more specifically addressing the first stages of biomineral maturation processes (Rey et al. 1995).

138 2. Materials and Methods

139 2.1. Synthesis of nanocrystalline apatite compounds

¹⁴⁰ Biomimetic (non-carbonated) nanocrystalline apatite compounds were prepared by precipitation from ¹⁴¹ mixing aqueous solutions of di-ammonium hydrogenphosphate (0.6 M) and calcium nitrate (0.3 M), at ¹⁴² 22 °C and at pH = 7.2 close to the physiological value. The excess of phosphate ions in solution, ¹⁴³ relative to the formation of hydroxyapatite, provides an internal pH buffer without any additives in the ¹⁴⁴ precipitating medium. After rapid mixing (1 min), the precipitates were left to mature (ageing in ¹⁴⁵ solution) for different periods of time, namely 20 min, 3 h, 1 day, 3 days, 5 days, 1 week and 3 weeks. ¹⁴⁶ Then the precipitates were filtered on Büchner funnel, thoroughly washed with deionized water and ¹⁴⁷ freeze-dried (freeze-dryer set to -80 °C, residual pressure 10 mbar).

Stoichiometric HA was prepared following a previously reported protocol (Raynaud et al. 2002). ¹⁴⁹ Briefly, the precipitation was carried out under reflux at 90 °C and pH = 8.5 from adding dropwise a ¹⁵⁰ solution of di-ammonium phosphate into a solution of calcium nitrate. The synthesis was carried out ¹⁵¹ under argon atmosphere to avoid atmospheric contamination, especially of CO₂. The reactants were ¹⁵² used in stoichiometric proportions, in the presence of ammonia for pH stabilization. The mixture was ¹⁵³ aged for 90 min prior to filtration on Büchner funnel and washing with deionized water. The ¹⁵⁴ precipitate was then oven-dried at 80 °C for 24 h and calcined at 1000 °C for 15 h.

 β -TCP was obtained by calcining at 900 °C for 16 h some amorphous calcium phosphate (am- $156 \text{ Ca}_3(\text{PO}_4)_2$) prepared by rapid precipitation from calcium nitrate (0.36 M) and di-ammonium 157 hydrogenphosphate (0.154 M) solutions under strongly alkaline conditions (pH = 10) after the protocol 158 proposed by Heughebaert and Montel (Heughebaert and Montel 1982).

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160 2.2. Characterization techniques

¹⁶¹ Powder X-ray diffraction (XRD) was performed for crystal structure identification, using a Brüker D8

¹⁶² Advance diffractometer with the monochromatic CuK_{α 1} radiation ($\lambda = 1.5406$ Å, step 0.021°). XRD ¹⁶³ profile fitting was performed using the JANA 2006 software. The crystallite mean length was ¹⁶⁴ estimated by applying the Hosemann and Vogel model (Vogel and Hosemann 1970) to the (002) and ¹⁶⁵ (004) planes, as this model takes into account the possible existence of non-negligible crystal disorder ¹⁶⁶ effects.

¹⁶⁷ Fourier transform infrared (FTIR) was used, in transmission mode, for complementary phase ¹⁶⁸ identification. The experiments were carried out on a Perkin Elmer 1600 spectrometer, in the ¹⁶⁹ wavenumber range 400-4000 cm⁻¹ and at 4 cm⁻¹ resolution.

The calcium content of the solids was determined by complexometry with EDTA (Charlot 1974). The relative error is 0.5%. Orthophosphate ionic contents (PO₄³⁻, HPO₄²⁻) were measured by traccolorimetry ($\lambda = 460$ nm) based on the yellow phospho-vanado-molybdenum complex VO₃[P(MO₃O₁₀)₄] formed in acidic conditions (Gee and Dietz 1953). Measurements were carried out tra in quartz holder with an UV-visible Hitachi Instruments U-1100 single beam spectrophotometer. These orthophosphate ion titrations have a relative error of 0.5 %. The amount of the protonated tra species HPO₄²⁻ ions is derived by comparing titrations carried out before and after calcining the tra samples at 600 °C for 1 h, which leads to the condensation of HPO₄²⁻ ions into pyrophosphate ions trac (P₂O₇⁴⁻) that do not form the yellow complex, as detailed by Gee and Dietz (Gee and Dietz 1953).

The thermal behavior of the differently matured apatites was followed by thermogravimetry (TG) analyses carried out in air on a Setaram SETSYS Evolution apparatus (heating rate 2.5 °C/min, temperature range 20 – 900 °C). Water contents of the starting powders were derived from the weight loss observed in the temperature range 20 – 300 °C.

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184 2.3. High Temperature Oxide Melt Solution Calorimetry

¹⁸⁵ High-temperature drop solution calorimetry was carried out in a Tian-Calvet twin calorimeter, as

¹⁸⁶ described in detail by Navrotsky (Navrotsky 1977, 1997). Drop solution enthalpies were measured by ¹⁸⁷ dropping 5 mg pressed pellets of material directly from room temperature, 298 K, into the molten ¹⁸⁸ solvent in the calorimeter, 973 K. Sodium molybdate (3Na₂O ⁻ 4MoO₃) was selected as appropriate ¹⁸⁹ solvent for this work based on earlier data (Ushakov et al. 2001) showing that all phosphorus was ¹⁹⁰ retained in the melt after dropping P₂O₅ or other phosphate-containing compounds.

Total calorimetric reaction times during calorimetry were in all cases less than 1 h. The shape of the ¹⁹² calorimetric peaks was consistent with rapid sample dissolution during the first few minutes of ¹⁹³ reaction. The end of the reaction was judged by the return of the baseline to its initial value. A ¹⁹⁴ minimum of 8 values were obtained for each composition, and uncertainties are two standard ¹⁹⁵ deviations from the mean value. During the experiments, air was flushed through the gas space above ¹⁹⁶ the melt (~80 mL/min) so as to accelerate the elimination of gases produced (H₂O or CO₂).

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198 3. Results and Discussion

199 3.1. Characterization

The calcium phosphate apatite samples prepared in this study with varying maturation times (**Table 1**) ²⁰⁰ were characterized by complementary techniques. TEM observations showed that all prepared samples ²⁰¹ exhibited a platelet morphology (see typical example of the 1-day maturation sample on **Figure 1**). ²⁰³ This plate-like morphology is characteristic of that of bone apatite (morphological biomimetism) as ²⁰⁴ reported in the literature (Johansen and Parks 1960). Powder XRD patterns exhibited diffraction peaks ²⁰⁵ that could all be attributed to a hydroxyapatite-like phase (hexagonal, P6₃/m space group, PDF card ²⁰⁶ 09-0432) as indicated in **Figure 2**. The samples are characterized by a low degree of crystallinity, as ²⁰⁷ for natural bone mineral (Gomez-Morales et al. 2013) or immature enamel crystals (Rey et al. 1995; ²⁰⁸ Gomez-Morales et al. 2013). The crystallinity of these synthetic materials progressively increases upon ²⁰⁹ maturation in accordance with previous data (Neuman et al. 1956; Vandecandelaere et al. 2013). This ²¹⁰ increased degree of crystallinity is indicated by the better resolution of the XRD patterns, which is ²¹¹ especially visible for peaks (002), (004) and (310) in the $2\theta = 28-36^{\circ}$ range. XRD peak broadening ²¹² analysis, using the Hosemann and Vogel model (Vogel and Hosemann 1970), led to the evaluation of ²¹³ mean crystallite dimensions. The longest mean crystallite dimension is accessible from analysis of the ²¹⁴ (002) and (004) peaks and increases from *ca*. 12 to 27 nm for maturation times ranging from 20 min to ²¹⁵ 3 weeks, thus confirming the nanocrystalline character of these samples.

FTIR analyses further confirm the apatitic nature of the samples (**Figure 3a**). Detailed observation of the spectra reveal, as expected from previous studies (Rey et al. 2007a; Rey et al. 2007b), the presence of non-apatitic contributions, especially in the v_4PO_4 vibration region (400-800 cm⁻¹) and more specifically at 535 cm⁻¹ (non-apatitic HPO₄²⁻ ions) and 617 cm⁻¹ (non-apatitic PO₄³⁻). For more mature samples, the libration band of apatitic OH⁻ at 631 cm⁻¹ becomes visible (see **Figure 3b**, illustrating an apatite matured for 1 week). The presence of non-apatitic contributions substantiates again the biomimetic character of these apatite compounds (Eichert et al. 2008), and it stresses their departure from the structure and composition of coarse hydroxyapatite "reference" material.

²²⁴ Calcium and phosphate titrations along with thermal analyses and the condition of electroneutrality ²²⁵ enabled the determination of the contents of each ionic species and water in every sample produced ²²⁶ with increasing maturation times. The determination of HPO₄/PO₄ relative amounts is made possible ²²⁷ for such apatite samples due to the absence of carbonate ions, therefore enabling one to determine ²²⁸ accurate chemical compositions needed for calorimetric evaluations. Indeed, the HPO₄/PO₄ balance is ²²⁹ generally drawn from orthophosphate titration, by comparing results obtained before and after heating ²²⁰ the samples at 600 °C (which decomposes HPO₄²⁻ into non-titrable pyrophosphates P₂O₇⁴⁻), while ²²¹ carbonate ions may interfere with these titration methods by partially reacting with HPO₄²⁻ ions ²²² through reactions such as CO₃²⁻ + 2HPO₄²⁻ \rightarrow CO₂ + 2PO₄³⁻ + H₂O (Eichert et al. 2008).

²³³ The water content has been assessed from the measured weight loss observed by thermogravimetric

²²⁴ analyses between 20 and 300 °C (see Supplementary Figure AR1) as this range corresponds to the ²²⁵ release of such associated water molecules from apatite nanocrystals (Banu 2005). The chemical ²²⁶ composition of the samples is reported in **Table 1**. The calcium and hydroxyl contents increase with ²²⁷ maturation time, while the HPO₄²⁻ content decreases. These variations produce a progressive evolution ²²⁸ of the chemical composition of the samples toward the stoichiometry of hydroxyapatite. This evolution ²²⁹ can also be monitored by following the Ca/P mole ratio, which increases here from 1.42 to 1.54 (\pm ²⁴⁰ 0.02). The amount of associated water molecules also decreases as ageing in solution progresses. ²⁴¹ These trends suggest the progressive disappearance of the hydrated non-apatitic surface layer from the ²⁴² nanocrystals.

Although the chemical formula $Ca_{10-x}(PO_4)_{6-x}(HPO_4)_x(OH)_{2-x}$ is often used to describe an onstoichiometric apatites, it shows some limitations when applied to the current analytical data, especially for short maturation times. In contrast the formula proposed by Kühl and Nebergall 1963, $Ca_{10-x-Z}(PO_4)_{6-x}(HPO_4)_x(OH)_{2-x-2Z}$, involving lower calcium and hydroxide contents, satisfactorily describes the overall chemical composition of the biomimetic apatite phases prepared in this work, and the values of Z are also indicated in **Table 1**. Z was indeed found to become significant for the most immature samples. Taking into account the presence of « n » moles of water per unit formula in the freeze-dried samples, the complete chemical formula thus becomes: $Ca_{10-x-Z}(PO_4)_{6-x}(HPO_4)_x(OH)_{2-x-2Z}$.

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253 3.2. Enthalpies of formation

The determination of the enthalpy of formation from the oxides of these apatites ($\Delta H_{f,oxides}$) from measured enthalpies of drop solution (ΔH_{ds}) requires the application of a thermodynamic cycle, indicated in **Table 2**. This cycle uses the experimental value of ΔH_{ds} measured for each hydrated apatite corresponding to various maturation stages, as well as the enthalpies of drop solution of ²⁵⁸ calcium carbonate CaCO₃ (calcite) and phosphorus oxide P₂O₅. The latter was determined previously ²⁵⁹ by Ushakov et al. 2001. Calcium carbonate was preferred to calcium oxide in this study taking into ²⁶⁰ account the difficulty to keep anhydrous CaO which has a tendency to partially transform into calcium ²⁶¹ hydroxide. The enthalpies of formation of the apatites from the elements, ΔH_f° , can then be calculated ²⁶² by adding the appropriate literature values of enthalpies of formation of the binary oxides from their ²⁶³ elements. The application of this cycle to reference compounds, namely β-tricalcium phosphate (β-²⁶⁴ TCP) and stoichiometric hydroxyapatite (**Table**), led to ΔH_f° values of -4090.2 ± 10.6 kJ/mol and -²⁶⁵ 13431.0 ± 22.7 kJ/mol respectively. These numbers are in good agreement (within ~ 0.7 %) with ²⁶⁶ values reported in the literature (Robie and Hemingway 1995) (-4120.8 ± 5.0 kJ/mol and -13477.0 ± ²⁶⁷ 10.0 kJ/mol respectively, the latter value being expressed for the unit formula Ca₁₀(PO₄)₆(OH)₂ rather ²⁶⁸ than Ca₅(PO₄)₃OH)), thus validating this cycle and measurements.

The application of the cycle then leads (**Table**) to the evaluation of the standard enthalpies of ²⁷⁰ formation of the apatite samples as prepared (hydrated) as well as those of the apatite phases ²⁷¹ themselves (anhydrous). As indicated in **Table 2**, the latter were obtained by considering hydration ²⁷² water molecules as thermodynamically equivalent to liquid water, as is often the case for hydrated ²⁷³ phases in which H₂O is not tightly bound and can be released below 300 °C (Drouet and Navrotsky ²⁷⁴ 2003).

The enthalpy values thus obtained, either relative to the elements or to the oxides, become more regative (exothermic) as maturation progresses. ΔH_f° varies from -12058.9 ± 12.2 to -12771.0 ± 21.4 kJ/mol for maturation times increasing from 20 min to 3 weeks (**Figure 4a**), thus approaching the value for stoichiometric HA, i.e. -13431.0 ± 22.7 kJ/mol. The $\Delta H_f^{\circ} = f(t)$ curve follows a monotonic, nearly exponential trend with faster changes during the first 3 days of ageing and slower progression beyond this stage. This trend is then found to parallel the evolutions (in the opposite direction) of the 280 Ca²⁺ and OH⁻ ion contents of the maturing apatite phases (see Supplementary Figure AR2). Based on 282 these findings, the enthalpy of formation of a nanocrystalline apatite phase appears to be directly related to its calcium and hydroxide contents. Indeed, the plot of ΔH_f° versus Ca²⁺ or OH⁻ ion content 284 shows roughly linear variations (see Supplementary Figure AR3), with fit parameters taking into account the cumulated experimental uncertainties ($R^2 = 0.886$ and 0.937 respectively). Since the ²⁸⁶ calcium content can be determined easily by techniques such as EDTA complexometry, ICP-AES or ²⁸⁷ atomic absorption spectroscopy, and is often reported in literature studies (as opposed to the hydroxide ²⁸⁸ content which is less easily accessible) we report here specifically the equation found for the linear fit ₂₈₉ obtained versus the apatite calcium content: $\Delta H_f^{\circ}(apatite) = -903.8 * [Ca^{2+} content] - 4426.7$, in $_{200}$ kJ/mol, and the relative error on ΔH_{f}° (apatite) can be estimated to be 1.0 % (see Figure 4b). These ²⁹¹ findings thus allow us to unveil, for the first time quantitatively, the direct correlation between the ²⁹² energetics of formation of biomimetic apatites and their ionic contents. Since the amount of calcium is ²⁹³ directly linked to the number of cationic vacancies, and similarly the hydroxide content to the amount $_{294}$ of anionic vacancies, ΔH_{f}° is found to fundamentally depend on the apatite maturation state: the ₂₉₅ system gets more energetically favorable (more exothermic ΔH_{f}°) as there are fewer crystal "defects" $_{296}$ in the structure. The linear variation of enthalpy with Ca²⁺ or OH⁻ contents also supports the fact that 207 this trend is rather independent on the distribution of the calcium ions between the surface hydrated ²⁹⁸ layer and the apatitic core: it thus allows one to extend the use of this prevision trend to other synthesis ²⁹⁹ scenarios leading to nanocrystalline apatites as final product. It should be noted that the contribution of ³⁰⁰ the non-apatitic chemical environments to the energetics cannot at present be separated from the ³⁰¹ overall energetic trend. Finally, these findings provide weak evidence that clustering or ordering of ³⁰² such defects does not occur with major energetic consequences.

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304 3.3. Entropies and Gibbs free energies

³⁰⁵ Entropy values for nonstoichiometric nanocrystalline apatites are not accessible from the literature.

³⁹⁵ Data reported for well-crystallized stoichiometric apatitic compounds such as hydroxy-, fluor- and ³⁹⁷ chlor-apatites are on the contrary available (Jemal et al. 1995; Jemal 2004; Ben Cherifa and Jemal ³⁹⁸ 2004), and a calculation reveals that, in all cases, the entropy contributions of $T \Delta S_f^{\circ}$ represent only a ³⁹⁹ minor proportion of the Gibbs free energy of formation $\Delta G_f^{\circ} = \Delta H_f^{\circ} - T \Delta S_f^{\circ}$ (of the order of 6 %, see ³¹⁰ Supplementary Figure AR4) compared to the enthalpy contribution ΔH_f° . Therefore the relative ³¹¹ stability of such compounds, which are theoretically assessed by comparing ΔG_f° values, can also be ³¹² reached in a more direct way by comparing the enthalpies of formation accessed by calorimetry. The ³¹³ data reported in **Figure 4a** therefore suggest that the relative stability of biomimetic apatites increases ³¹⁴ as maturation progresses, evolving toward the level of stoichiometric hydroxyapatite (without reaching ³¹⁵ it though).

³¹⁶ We obtain a better estimate of the entropy of nanocrystalline apatites by considering the following ³¹⁷ reaction:

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$$(9-x) \operatorname{CaO}_{(s, 298)} + 3 \operatorname{P_2O}_{5(s, 298)} + (1-Z) \operatorname{Ca}(OH)_{2(s, 298)} \rightarrow \operatorname{Ca}_{10-x-Z}(\operatorname{PO}_4)_{6-x}(\operatorname{HPO}_4)_x(OH)_{2-x-2Z(s, 298)}$$

$$(Eq. 1)$$

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Since this reaction only involves solid phases, the corresponding entropy change, ΔS°_{react} , is expected to be close to zero. Considering $\Delta S^{\circ}_{react} = 0$, T = 298 K and entropy values for CaO, P₂O₅ and Ca(OH)₂ from thermodynamic databases (Robie and Hemingway 1995), this reaction leads for stoichiometric HA (x = Z = 0) to the standard entropy S°(HA) = 769.5 J/(mol.K). This value is in reasonable agreement (within 1.5 %) with the value 780.8 J/(mol.K) reported in the literature (Robie and Hemingway 1995) (expressed for the unit formula Ca₁₀(PO₄)₆(OH)₂) thus supporting this estimation method. In a similar way, the entropy S° of each nanocrystalline apatite from this work was evaluated, as well as the corresponding standard entropy of formation from the elements ΔS_{f}° (**Table** ³³⁰ **4**). The Gibbs free energies of formation ΔG_f° could then also be derived, at 298 K, from the ΔH_f° and ³³¹ ΔS_f° values. These data show again that the entropy contributions to the ΔG_f° values are small ³³² compared to enthalpy contributions, and that the Gibbs free energy becomes more negative (favorable) ³³³ as the system gets more mature (**Figure 4a**). This conclusion probably still holds even if one adds the ³³⁴ contribution of a possible configurational entropy resulting from the location of defects in the ³³⁵ structure, but this contribution cannot be calculated accurately. Note that entropy contributions may ³³⁶ presumably also be approximated using computational methods. However, such calculations would ³³⁷ require good knowledge of stuctural features, while the exact location of ions contained in the hydrated ³³⁸ layer on such biomimetic apatites is still essentially undertermined.

³³⁹ A major variation of ΔG_f° similar to that in ΔH_f° is found during the first days of maturation in ³⁴⁰ solution, while the decrease in ΔG_f° becomes less pronounced beyond a few days. A plot of ΔG_f° ³⁴¹ versus calcium content again leads to a linear trend (see Supplementary Figure AR3), corresponding to ³⁴² the equation ΔG_f° (apatite) = -843.6 * [Ca²⁺ content] – 4204.5 (in kJ/mol, R² = 0.880, relative error ³⁴³ estimated to 1.1 %). This equation then allows one to draw predictive estimates of the value of ΔG_f° ³⁴⁴ for other calcium phosphate nanocrystalline apatites, based on the determination of their calcium ³⁴⁵ content.

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347 3.4. Apatite maturation energetics

It is desirable to estimate the variation in free energy of maturation $\Delta G_{maturation}(i \rightarrow f)$, corresponding to the maturation process in solution transforming an apatite from an initial maturation stage « i » to a more advanced stage « f ». To this aim, it is necessary to take into account the thermodynamic data for the aqueous ions incorporated or released during this maturation process in solution. This task is however more difficult as it may appear at first sight. Indeed, the maturation process is a complex phenomenon where not only the chemical composition evolves towards stoichiometry, but also where ³⁵⁴ secondary reactions such as dissolution-reprecipitation may play a role. However, the global change in ³⁵⁵ chemical composition during maturation can probably be considered as a dominant phenomenon, since ³⁵⁶ the overall ion content can significantly change upon maturation as illustrated by **Table 1**, which is ³⁵⁷ bound to quantitatively impact the compounds thermodynamic properties.

Considering the simplified scenario where only the global change in apatite composition is taken 358 into account, it is possible to determine in particular the sign of $\Delta G_{\text{maturation}}(i \rightarrow f)$ based on a modeled ³⁶⁰ maturation reaction. The increase in Ca/P ratio that accompanies the observed evolution toward ³⁶¹ stoichiometry could theoretically either be explained by an additional incorporation of Ca²⁺ ions ³⁶² (increase of numerator) or by a release of phosphate ions in the medium (decrease of denominator), or ³⁶³ both. However, the concentration of free calcium ions in solution is likely to be extremely low due to ³⁶⁴ the large excess of phosphate ions in the synthesis medium or to the presence of numerous calcium-³⁶⁵ complexing entities in body fluids *in vivo* (phosphates, carbonates and protein ionic species). $_{366}$ Therefore, the possibility to incorporate additional Ca²⁺ ions from the solution appears unlikely. In ³⁶⁷ contrast, the release of phosphate ions from the solid to the solution appears much more probable, ³⁶⁸ especially as a protonated form which is stable under physiological pH. Since phosphate ions from the ³⁶⁹ non-apatitic surface layer are mostly protonated as HPO_4^{2-} , while the amount of HPO_4^{2-} in the solid ³⁷⁰ decreases upon maturation (in both synthetic and biological apatites) (Rey et al. 1991b; Cazalbou et al. $_{371}$ 2004a) in favor of non-protonated PO₄³⁻, the release of phosphate as H₂PO₄⁻ appears as the most ³⁷² probable route, which may be described by the following scheme, involving proton hopping between $_{373}$ two surface HPO₄²⁻ ions:

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$$2 \text{HPO}_4^{2^-}(\text{solid}) \rightarrow \text{H}_2\text{PO}_4^{-}(\text{released in the solution}) + \text{PO}_4^{3^-}(\text{solid})$$
 (Eq. 2)

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 $_{377}$ Since this departure of anionic $H_2PO_4^-$ ions from the solid would lead to a decrease in negative

³⁷⁸ charges, it has to be compensated by a simultaneous incorporation of OH⁻ ions. This was indeed found ³⁷⁹ experimentally by the increased hydroxylation of apatites upon maturation (**Table 1**). In this context, ³⁸⁰ the global reaction scheme describing the change in composition during the maturation process may ³⁸¹ probably be written as:

382

Apatite in state
$$(i * i) + \delta_1 H_2 O_{(liq)} \rightarrow Apatite in state (f') + \delta_2 H_2 PO_4(aq) + \delta_3 H^+(aq)$$
 (Eq. 3)

384

³⁸⁵ where the chemical species $H_2O_{(liq)}$ and $H^+(aq)$ have been preferred to the direct involvement of $OH^-_{(aq)}$ ³⁸⁶ ions, due to the neutral pH where such maturations were carried out.

The variation in Gibbs free energy accompanying this reaction can be written as $\Delta G_{maturation}(i \rightarrow f) = \Delta G^{\circ}_{maturation}(i \rightarrow f) + RT * ln(K)$ where K, the equilibrium constant, is given by the activity product: ³⁸⁷ (H₂PO₄⁻(aq))⁸² * (H⁺(aq))⁸³. Based on the data in Table 4 and on thermodynamic data (Wagman et al. ³⁸⁹ 1982; Robie and Hemingway 1995) for H₂O_(liq) and H₂PO₄⁻(aq), the values of $\Delta G^{\circ}_{maturation}(i \rightarrow f)$ were ³⁹¹ calculated at 298 K for various maturation stages « f » relative to the maturation of 20 min taken as ³⁹² reference (initial state « i »). Also, under physiological conditions (considering pH = 7.4 and (H₂PO₄⁻) ³⁹³ $\cong 10^{-4}$ M), the values of the RT * ln(K) term for each sample were determined. The obtained values of ³⁹⁴ $\Delta G_{maturation}$ (at 298 K) are plotted in **Figure 5**. The dispersion of the points is probably linked to the ³⁹⁵ simplistic scenario considered here, not taking into account secondary surface reaction in particular. ³⁹⁶ Interestingly, the value of $\Delta G_{maturation}$ is found to be negative in all cases, ranging from 0 to -117 ± 23 ³⁹⁷ kJ/mol, with the most negative value corresponding to evolution toward coarse stoichiometric HA ³⁹⁸ (corresponding to -185 ± 15 kJ/mol). These findings give a quantitative background for biomimetic ³⁹⁹ calcium phosphate apatite ageing (studied here on synthetic samples over a period of 3 weeks ⁴⁰⁰ maturation).

The spontaneous character of this maturation process can thus be considered as a thermodynamic

driving force explaining the inexorable evolution of nonstoichiometric apatite nanocrystals (e.g. found in immature enamel and in young bone) toward more stable states. Such states are characterized by 1) are a composition closer to stoichiometry and 2) an associated decrease in surface reactivity and solubility. This stabilization (in terms of both thermodynamics and kinetics) is advantageous in the case of enamel maturation, taking into account the final functions of this biomineral *in vivo* (i.e. resistance to external aggressions of various nature including chemical). On the contrary, it is not beneficial in the are case of bone which needs to remain relatively soluble and reactive (i.e. able to exchange ions with surrounding fluids, and/or to undergo dissolution/reprecipitation phenomena upon remodeling). Our findings thus suggest that bone remodeling has a strong thermodynamic basis. Indeed, according to our the data, the metastable apatite composing newly-formed bone matter is thermodynamically driven to are inevitably transform into a more stable, less soluble and less reactive state, with lower surface area, minimized non-apatitic surface layer, and fewer reactive surface sites. Such lowered surface reactivity twas for example observed in model experiments run on synthetic nanocrystalline apatite and on are chicken bone (Cazalbou et al. 2004a).

Thus the maturation of bone apatite crystals with time is bound to lead to limited capability in body 417 fluids homeostasis. Therefore, the above-quantified thermodynamic *driving force* (negative 418 $\Delta G_{maturation}$) along which reactive but immature apatite nanocrystals evolve toward more mature but 419 less reactive states could be seen as a physical-chemical (rather than purely biological) basis 420 explaining the need for bone to be regularly remodeled. Then, such remodeling does not only allow 421 skeletal growth from infancy to adulthood as well as self-repair after bone injury (healing of bone 422 tissue microfractures linked to pathological or traumatic events), but it is a "necessity" in view of 423 conserving highly-reactive bone biomineral crystals capable of playing their role in homeostasis (e.g. 424 as a participation in the stabilization of calcium, magnesium, strontium, phosphate concentrations in 425 body fluids). Bone remodeling, which "resets" the maturation process, at the biological cost of the 426 energy and nutrients required for it, is thus favorable and necessary to the organism.

⁴²⁷ In addition to enabling the estimation of $\Delta G_{maturation}$, the determination of free energies of formation ⁴²⁸ ΔG_f° for such apatite compounds can be used for other thermodynamic calculations. One obvious other ⁴²⁹ example concerns the evaluation of the solubility product of such biomimetic apatites as a function of ⁴³⁰ their maturation state. The question of solubility is indeed relevant when dealing with enamel ⁴³¹ formation or bone remodeling processes.. In the case of hydroxyapatite, the dissolution equilibrium ⁴³² can be described by the reaction:

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$$Ca_{10}(PO_4)_6(OH)_2 + 2 H^+_{(aq)} \leftrightarrow 10 Ca^{2+}_{(aq)} + 6 PO_4^{3-}_{(aq)} + 2 H_2O_{(liq)}$$
 (Eq. 4)

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⁴³⁶ The variation of free energy ΔG°_{disso} accompanying this reaction is linked to the solubility product K_{sp} ⁴³⁷ by the equation:

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$$\Delta G^{\circ}_{disso} = -2.303 * RT * log(K_{sp}) = 2.303 * RT * pK_{sp}$$
 (Eq. 5)

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⁴⁴¹ Considering the experimental compositions found in this work, the strict application of this equation ⁴⁴² at 298 K leads to pK_{sp} values ranging from 96 to 113 (see details in Supplementary Figure AR5). As a ⁴⁴³ general tendency, these calculations suggest that apatite solubility decreases as maturation progresses, ⁴⁴⁴ tending toward the value for stoichiometric HA (McDowell et al. 1977) ($pK_{sp}(HA) = 117$). However, ⁴⁴⁵ several literature studies (Hsu et al. 1994; Baig et al. 1996; Chhettry et al. 1999) have pointed out ⁴⁴⁶ noticeable differences between the *apparent* solubility of nonstoichiometric calcium phosphate apatites ⁴⁴⁷ and their *theoretical* value, despite long periods of stabilization in solution, thus showing that the ⁴⁴⁸ system did not reach the true thermodynamic equilibrium. This phenomenon depicts a situation where ⁴⁴⁹ dissolution (relatively rapid at first) has essentially stopped and where nucleation/growth processes are ⁴⁵⁰ not discernable, over the temporal scale of the experiments. Also the non-constancy of the solubility ⁴⁵¹ product for such compounds was unveiled by these studies, as it was found to depend on the fraction ⁴⁵² of mineral dissolved. This behavior, referred to as Metastable Equilibrium of Solubility (MES), was ⁴⁵³ then found to be related to the existence of microstrains (Higuchi et al. 1984) within the constitutive ⁴⁵⁴ non-ideal crystals. Nanocrystalline biological and biomimetic apatites exhibit a non-homogeneous ⁴⁵⁵ chemical composition as the nanocrystals are constituted of an apatitic core surrounded by a non-⁴⁵⁶ apatitic surface layer. In these conditions, the observation of altered apparent solubilities – as ⁴⁵⁷ compared to theoretical values – is rather unsurprising. Due to the incongruence of the dissolution of ⁴⁵⁸ such nanocrystalline compounds, it should be kept in mind that such pK_{sp} values drawn from ΔG°_{disso} ⁴⁵⁹ may only be considered for pointing out the decreasing solubility of biomimetic apatites upon ageing. ⁴⁶⁰ These pK_{sp} values should however be considered with caution for determining precise calcium and ⁴⁶¹ phosphate concentrations in surrounding solutions: for this purpose, experimental solubility tests ⁴⁶² remain the best approach.

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464 3.5. Implications

The inexorable evolution of immature apatite crystals with time is advantageous in the case of enamel where increased stability and lower solubility are beneficial to the "protection" functionality of this biomineral. In contrast, a similar thermodynamically-driven evolution of apatite is deleterious for bone mineral which plays a key role in the regulation of body fluid ionic concentrations *in vivo* through homeostasis. These results strongly suggest that bone remodeling could be seen as a thermodynamic necessity to eliminate the "too stable" and poorly bioactive aged apatite crystals in favor of the neo-formation of immature, less stable, and highly reactive nanocrystals. The timetra dependent evolution of apatite-based calcified tissues such as bone or enamel could thus be dictated – tra at least in great part – by a thermodynamic driving force, despite the current emphasis on a mostly ⁴⁷⁴ biologically driven process. Our data strongly support this new interpretation, with a major role played ⁴⁷⁵ by mineral thermodynamics.

⁴⁷⁶ Beside implications of these thermodynamic data for solubility behavior of nanocrystalline ⁴⁷⁷ biomimetic apatites, these findings are bound to find other implications. In the domain of ectopic ⁴⁷⁸ (abnormal or untypical) calcifications, several mineral compounds may be observed *in vivo* (e.g. ⁴⁷⁹ apatite, pyrophosphate, whitlockite, struvite). Specific reasons for their formation are not yet clearly ⁴⁸⁰ determined. Yet, thermodynamic stabilities are probably involved in the persistence or transformations ⁴⁸¹ of such ectopic mineralizations. Biomaterials for bone replacement based on nanocrystalline apatites ⁴⁸² show great promise due to their high surface reactivity. However any synthesis or post-synthesis step ⁴⁸³ (e.g. sterilization) which may involve humid conditions and/or heating should be considered cautiously ⁴⁸⁴ and with good understanding of nanocrystallie apatite physic-chemistry, since further evolution of the ⁴⁸⁵ calcium phosphate systems may prove useful in other domains such as (geo)microbiology (e.g. ⁴⁸⁷ calcifications occurring inside eukaryotic cells) (Raven and Knoll 2010) or link to the evolution of life ⁴⁸⁶ on Earth (e.g. evolution from carbonate-based shells to phosphate-based skeletons).

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633 Figure captions:

635 636 637 Figure 1: TEM micrograph for a biomimetic apatite sample matured for 1 day 638 639 Figure 2: XRD pattern for nanocrystalline apatite samples with varying maturation times 640 641 Figure 3: a) FTIR spectrum for a nanocrystalline apatite sample matured 1 day, b) detail in the 4PO4 vibration region for a sample 642 matured 1 week 643 ⁶⁴⁴ Figure 4: a) Evolution of ΔH_f° and ΔG_f° for nanocrystalline apatites versus maturation time in solution (uncertainties are two standard ⁶⁴⁵ deviations, SD, of the mean), and b) linear fit for $\Delta H_f^{\circ} = f(Ca^{2+} \text{ content})$ Figure 5: Evolution of $\Delta G_{maturation}(i \rightarrow f)$ versus maturation time, taking the 20-min-matured sample as initial state 647 648 649 650 651 652 653 654 Table captions: Table 1: Chemical composition of apatite samples (estimated uncertainty on each ion content: 0.5%) 657 658 Table 2: Thermodynamic cycle used in the calculations of $\Delta H_{foxides}$ and ΔH_{f}° 659 660 **Table 3:** Experimental ΔH_{ds} values and derived $\Delta H_{foxides}$ and ΔH_{f}° for nanocrystalline apatites and for reference compounds HA and β -661 662 TCP 663 **Table 4:** Evaluation of S°, ΔS_f° and ΔG_f° for nanocrystalline apatites matured between 20 min and 3 weeks and for stoichiometric HA 664 665

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668 Figure 1: TEM micrograph for a biomimetic apatite sample matured for 1 day



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673 Figure 2: XRD pattern for nanocrystalline apatite samples with varying maturation times

Figure 3: a) FTIR spectrum for a nanocrystalline apatite sample matured 1 day, b) detail in the v_4PO_4 vibration region for a sample matured 1 week

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⁶⁹⁷ Figure 5: Evolution of $\Delta G_{\text{maturation}}(i \rightarrow f)$ versus maturation time, taking the 20-min-matured sample as initial state (± SD)



699 Table 1: Chemical composition of apatite samples (estimated uncertainty on each ion content: 0.5%)

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Maturation	Ca/P	Value	Contents (moles per apatite unit formula *)				
time	mole ratio	of Z *	Ca ²⁺	PO ₄ ³⁻	HPO ₄ ²⁻	OH-	H ₂ O
20 min	1.43	0.42	8.55	4.97	1.03	0.13	5.94
3 h	1.42	0.43	8.54	4.97	1.03	0.12	4.18
1 d	1.44	0.24	8.64	4.88	1.12	0.39	3.60
3 d	1.48	0.39	8.86	5.25	0.75	0.46	3.53
5 d	1.49	0.12	8.92	5.04	0.96	0.80	3.21
1 wk	1.49	0.17	8.94	5.11	0.89	0.77	2.86
3 wk	1.54	0.07	9.21	5.28	0.72	1.15	3.28

⁷⁰¹ * considering Kühl and Nebergall's expression Ca_{10-x-Z}(PO₄)_{6-x}(HPO₄)_x(OH)_{2-x-2Z}

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 $_{705}$ Table 2: Thermodynamic cycle used in the calculations of $\Delta H_{f}^{\,\circ}$

Reacti	ons:	ΔH
(1)	$Ca_{10-x-Z}(PO_{4})_{6-x}(HPO_{4})_{x}(OH)_{2-x-2Z}(H_{2}O)_{n} \rightarrow (10-x-Z) CaO (soln, 973) + 3 P_{2}O_{5} (soln, 973) + (1-Z+n) H_{2}O (g, 973) + $	$\Delta H_{ds}(apatite, hydrated)$
	Reactions from oxides:	
(2)	$CaCO_3 (s, 298) \rightarrow CaO (soln, 973) + CO_2 (g, 973)$	$\Delta H_{ds}(CaCO_3)$
3)	$CO_2(g, 298) \to CO_2(g, 973)$	$\Delta H_{hc}(CO_2(g))$
4)	$P_2O_5(s, 298) \rightarrow P_2O_5(soln, 973)$	$\Delta H_{ds}(P_2O_5)$
5)	$H_2O(g, 298) \to H_2O(g, 973)$	$\Delta H_{hc}(H_2O(g))$
6)	$H_2O(1, 298) \rightarrow H_2O(g, 298)$	$\Delta H^{\circ}_{vap, 298}(H_2O(l))$
	Reactions from elements:	
7)	Ca (s, 298) + C (s, 298) + $3/2$ O ₂ (g, 298) \rightarrow CaCO ₃ (s, 298)	$\Delta H_{f}^{\circ}(CaCO_{3})$
8)	$C(s, 298) + O_2(g, 298) \rightarrow CO_2(g, 298)$	$\Delta H_{f}^{\circ}(CO_{2}(g))$
9)	$Ca (s, 298) + 1/2 O_2 (g, 298) \rightarrow CaO (s, 298)$	$\Delta H_{f}^{\circ}(CaO)$
10)	$2 P (s, 298) + 5/2 O_2 (g, 298) \rightarrow P_2O_5 (s, 298)$	$\Delta H_{f}^{o}(P_{2}O_{5})$
11)	$H_2(g, 298) + \frac{1}{2}O_2(g, 298) \rightarrow H_2O(g, 298)$	$\Delta H_{f}^{o}(H_{2}O(g))$
12)	$(10-x-Z) \text{ CaO} (s, 298) + 3 \text{ P}_{2}\text{O}_{5} (s, 298) + (2-x-2Z+n) \text{ H}_{2}\text{O} (l, 298) \rightarrow \text{ Ca}_{10-x-Z}(\text{PO}_{4})_{6-x}(\text{HPO}_{4})_{x}(\text{OH})_{2-x-2Z}(\text{H}_{2}\text{O})_{n}$	$\Delta H_{f, oxides}^{\circ}$ (apatite)
Theref	fora	
	$_{\text{ides}}^{\circ}$ (apatite) = $-\Delta H_1 + (10\text{-}x\text{-}Z) \Delta H_{\text{ds}}(\text{CaO}) + 3 \Delta H_4 + (2\text{-}x\text{-}2Z\text{+}n) \Delta H_5 + (2\text{-}x\text{-}2Z\text{+}n) \Delta H_6$	
	$= -\Delta H_1 + (10 \text{ x-}Z) [\Delta H_2 + \Delta H_7 - \Delta H_9 - \Delta H_8 - \Delta H_3] + 3 \Delta H_4 + (2 \text{ x-}2Z + n) \Delta H_5 + (2 \text{ x-}2Z + n) \Delta H_6$	
Form	ation of nanocrystalline apatites from the elements:	
(13)	$(10-x-Z)$ Ca (s, 298) + 6 P (s, 298) + $(1-Z+n)$ H ₂ (g, 298) + $(26-x-2Z+n)/2$ O ₂ (g, 298) \rightarrow	
	Ca _{10-x-Z} (PO ₄) _{6-x} (HPO ₄) _x (OH) _{2-x-2Z} (H ₂ O) _n	ΔH_{f}° (apatite, hydrated
Theref		
∆H _f ° (apatite, hydrated) = $-\Delta H_1 + (10-x-Z) \Delta H_2 + (10-x-Z) \Delta H_7 - (10-x-Z) \Delta H_3 - (10-x-Z) \Delta H_8 + 3 \Delta H_4 + 3 \Delta H_{10} + (1-Z+r_{10}) \Delta H_{10} + (1-$	h) $\Delta H_5 + (1-Z+n) \Delta H_{11}$

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711 Table 3: Experimental ΔH_{ds} values and derived $\Delta H_{f_{5}oxides}$ and ΔH_{f}° for nanocrystalline apatites and for reference compounds HA and β -712 TCP (\pm SD)

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Sample	$\frac{\Delta H_{ds}}{(kJ/mol)}$	ΔH _f ° (compound) (kJ/mol)	200 1/	1 of m
Reference com	pounds:		290 N	, 1 atm
β-ΤСΡ	267.3 ± 9.7 (6)*	-4090.2 ± 10.6		
HA stoich.	$1027.7 \pm 21.4 (11)$	-13431.0 ± 22.7		
Sample	ΔH _{ds} (kJ/mol)	ΔH _f ° (apatite, hydrated) (kJ/mol)	ΔH _{f oxides} (kJ/mol)	$\Delta {{H_{\rm f}}^\circ}\ (kJ/mol)$
Nanocrystallin	e apatites:			
20 min	$1197.7 \pm 10.0 (10)$	-13756.8 ± 12.2	-1952.2 ± 12.5	-12058.9 ± 12.2
3 hour	1198.2 ± 15.0 (9)	-13370.7 ± 16.5	-2073.9 ± 16.8	-12174.9 ± 16.5
1 day	1241.7 ± 9.5 (8)	-13393.4 ± 11.8	-2152.7 ± 12.1	-12364.4 ± 11.8
3 days	1088.6 ± 9.1 (9)	-13352.3 ± 11.5	-2032.8 ± 11.8	-12342.1 ± 11.5
5 days	1077.4 ± 5.1 (9)	-13373.3 ± 8.7	-2030.5 ± 9.2	-12457.0 ± 8.7
1 week	1137.2 ± 9.1 (10)	-13362.2 ± 11.5	-2119.5 ± 11.9	-12546.1 ± 11.5
3 weeks	1172.8 ± 20.2 (9)	-13708.7 ± 21.4	-2141.3 ± 21.6	-12771.0 ± 21.4

* numbers in parentheses indicate the number of drop solution calorimetry experiments

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⁷¹⁹ Table 4: Evaluation of S°, ΔS_f° and ΔG_f° for nanocrystalline apatites matured between 20 min and 3 weeks, and for stoichiometric HA (± ⁷²⁰ SD)

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	At 298 K			
Sample	Estimated S° J/(mol.K) [± 20 J/(mol.K)]	Estimated ΔS°_{f} J/(mol.K) [± 20 J/(mol.K)]	Recall of ΔH° _f kJ/mol	Corresponding ΔG° _f kJ/mol
Nanocrystalline apatite c	ompounds:			
20 min	695.2	-2 469	$-12\ 058.9 \pm 12.2$	$-11\ 323.1\pm12.2$
3 hour	694.6	-2 467	$-12\ 174.9 \pm 16.5$	$-11\ 439.7 \pm 16.5$
1 day	706.6	-2 512	$-12\ 364.4 \pm 11.8$	$-11\ 616.0\pm11.8$
3 days	708.2	-2 507	$-12\ 342.1 \pm 11.5$	-11595.0 ± 11.5
5 days	722.9	-2 565	$-12\ 457.0\pm 8.7$	$-11\ 692.6\pm 8.7$
1 week	721.4	-2 558	-12546.1 ± 11.5	$-11\ 783.9 \pm 11.5$
3 weeks	736.5	-2 607	$-12\ 771.0 \pm 21.4$	-11994.2 ± 21.4
Stoichiometric HA	769.5	-2 704	$-13\ 477\pm 10$	$-12\ 674.2\pm 10$

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728 Supplementary files as Additional Resources for the Review process:

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⁷³¹ Figure AR1: TG analysis on nanocrystalline apatite. Graph: example for 20-min maturation time. Inlet: evolutions of weight ⁷³² losses Δm_1 , Δm_2 and Δm_3 (as described below) versus maturation (SEM on Δm_1 : 0.01%, SEM on Δm_2 and Δm_3 : 0.1%)

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TG analysis – apatite matured for 20 min



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⁷⁴¹ Figure AR3: Plots of ΔH_f° versus Ca²⁺ (plot a) and OH⁻ (plot b) ion content in apatite, and of ΔG_f° versus Ca²⁺ (plot c), and ⁷⁴² related linear fits



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750 Figure AR4: Thermodynamic literature data for stoichiometric hydroxy-, fluor- and chlor-apatites (after Jemal et al. 1995; 751 Jemal 2004; Ben Cherifa et al. 2004)

	$\Delta G_{f 298}^{o}$ (kJ/mol)	$\Delta H_{f^{\circ}298} (kJ/mol)$	$\Delta S_{f^{\circ}298} (kJ/(mol.K))$
hydroxyapatites			
Ca10(PO4)6(OH)2	-12 674	-13 477	-2.69
Sr10(PO4)6(OH)2	-12 587	-13 373	-2.64
Pb10(PO4)6(OH)2	-7482	-8 261	-2.61
Cd10(PO4)6(OH)2	-7 873	-8 652	-2.61
Ba10(PO4)6(OH)2	-12 553	-13 309	-2.54
luorapatites			
Ca10(PO4)6F2	-12 781	-13 558	-2.61
Cd10(PO4)6F2	-8 045	-8 795	-2.52
Sr10(PO4)6F2	-12 845	-13 604	-2.55
Pb10(PO4)6F2	-7 782	-8 529	-2.51
Ba10(PO4)6F2	-12 834	-13 564	-2.45
chlorapatites			
Ca10(PO4)6Cl2	-12 418	-13 180	-2.56
Cd10(PO4)6Cl2	-7 719	-8 463	-2.50
Sr10(PO4)6Cl2	-12 478	-13 233	-2.53
Pb10(PO4)6Cl2	-7 458	-8 220	-2.56
Ba10(PO4)6Cl2	-12 418	-13 246	-2.78

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761 Figure AR5: Estimation of theoretical solubility products (at 298 K) for nanocrystalline apatites matured for varying periods of 762 time, as drawn from ΔG°_{disso} calculation 763 764 765 160 150 140 130 120 рК_{sp} ₫ 110 ◙ Ŧ 100 . y = 0.0002 x + 101.7 90 $(R^2 = 0.1283)$ 80 70 60 0 10000 20000 30000 Maturation time (min) 766 767

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